1	DYSREGULATED PLACENTAL MICRORNAS IN EARLY AND LATE ONSET PREECLAMPSIA
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23 CONFLICT OF INTEREST

- 24 The authors declare that they have no conflict of interest .
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27 ABSTRACT

- *Introduction:* To determine the miRNA expression profile in placentas complicated by Preeclampsia (PE)
 and compare it to uncomplicated pregnancies.
- 30 *Methods:* Sixteen placentas from women with PE, [11 with early onset PE (EOPE) and 5 with late onset PE
- 31 (LOPE)], as well as 8 placentas from uncomplicated pregnancies were analyzed using miRNA microarrays.
- 32 For statistical analyses the MATLAB® simulation environment was applied. The over-expression of miR-
- 33 518a-5p was verified using Quantitative Real-Time Polymerase Chain Reaction.
- 34 Results: Forty four miRNAs were found dysregulated in PE complicated placentas. Statistical analysis
- 35 revealed that miR-431, miR-518a-5p and miR-124* were over-expressed in EOPE complicated placentas as
- 36 compared to controls, whereas miR-544 and miR-3942 were down-regulated in EOPE. When comparing the
- 37 miRNA expression profile in cases with PE and PE- growth restricted fetuses (FGR), miR-431 and miR-
- 38 518a-5p were found over-expressed in pregnancies complicated by FGR.
- 39 Discussion: Since specific miRNAs can differentiate EOPE and LOPE from uncomplicated placentas, they
- 40 may be considered as putative PE-specific biomarkers. MiR-518a-5p emerged as a potential diagnostic
- 41 indicator for EOPE cases as well as for PE-FGR complicated placentas, indicating a potential link to the
- 42 severity of the disease.
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- 44 Keywords: miRNAs, placenta, microarrays, early onset Preeclampsia, late onset Preeclampsia
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47 ABBREVIATIONS

ABBREVIATION	EXPLANATION				
ADAM	A Disintegrin And Metalloproteinase				
AUC	Area Under the Curve				
CDK6	Cyclin Dependent Kinase 6				
cDNA	Complementary Deoxyribonucleic Acid				
EOPE	Early Onset PE				
FDR	False Discovery Rate				
FGR	Fetal Growth Restricted Fetuses				
IGF1R	Insulin Like Growth Factor 1 Receptor				
IUGR	Intrauterine Growth Restriction				
LOPE	Late Onset PE				
MAPK	Mitogen-Activated Protein Kinases				
miRNAs	MicroRNAs				
MMP	Matrix Metalloproteinase				
PE	Preeclampsia				
PTEN	Phosphatase and Tensin homolog				
PU	Proteinuria				
qRT-PCR	Quantitative Real-Time Polymerase Chain Reaction				
ROC curve	Receiver Operating Characteristic curve				
RUNX2	Runt-Related Transcription Factor 2				
TGF-β	Transforming Growth Factor beta				
TNF	Tumor Necrosis Factor				
TP53	Tumor Protein 53				
VCAM1	Vascular Cell Adhesion Molecule 1				
VEGFA	Vascular Endothelial Growth Factor A				

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51 INTRODUCTION

52 Preeclampsia (PE) is an obstetric complication affecting 2-8% of all pregnancies responsible for maternal, 53 fetal and neonatal morbidity and mortality worldwide [1]. Increasing evidence suggests that both women 54 who develop PE and their neonates have a higher risk of developing cardiovascular disease and metabolic 55 syndromes including type II diabetes, later in life. Traditionally, PE is characterized by new onset 56 hypertension and proteinuria after 20 weeks of gestation. Depending on the gestational age by which the symptoms develop, PE is classified as early-onset (EOPE-before 34 weeks of gestation) or late-onset (LOPE-57 at or after 34 weeks of gestation). Despite extensive efforts, the pathogenesis of the disease is not fully 58 59 understood. EOPE shows a greater association with impaired trophoblast invasion, ischemia at the placenta and fetal growth restriction (FGR) as compared to LOPE, whereas LOPE is not often associated with 60 61 placenta hypoperfusion and profound FGR [2].

MicroRNAs (miRNAs) belong to a highly conserved class of short non-coding molecules that regulate gene expression at post-transcriptional level [3]. It is estimated that 30% of human genes are transcriptionally negatively regulated by miRNAs, highlighting their crucial role in several cellular processes including cell growth, proliferation, differentiation and apoptosis [4]. Given the central role of the placenta in the development of PE, recent evidence suggests that miRNAs may be implicated in the pathogenesis of the disease regulating common pathways such as angiogenesis, hypoxia, ischemia and metabolism [5-14].

In the present study, we compared the placenta miRNA expression profiles of PE with those of uncomplicated pregnancies using a microarray hybridization based technology. The obtained miRNA patterns were correlated with clinical parameters known to be associated with PE including proteinuria, gestational age of disease onset and the presence of FGR fetus.

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74 PATIENTS AND METHODS

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76 Patient and tissue samples

77 Placental samples were collected from 16 women with PE (n=11 with EOPE and n=5 diagnosed with LOPE) 78 and from 8 with uncomplicated term pregnancies at the time of cesarean section in the 3rd Department of 79 Obstetrics and Gynecology at National and Kapodistrian University of Athens. Among the PE complicated cases, 8 carried a FGR fetus (PE-FGR): in the EOPE group, 6 out of 11 neonates had FGR confirmed by 80 81 birth weight percentile below the 10th whereas in the LOPE group 1 neonate was confirmed as FGR by birth 82 weight below the 5th percentile and the other was classified as FGR on the basis of a noted reduction of growth expressed by a static growth of abdominal velocity and abnormal placental Dopplers. Clinical 83 84 characteristics of women participating in the study are summarized in **Table 1**.

For each case, two pieces of placental tissue approximately 1×1×1cm in size were randomly obtained from the central part of the placenta maternal phase, as previously described [9]. Tissues were washed with isotonic saline solution, to remove excess of blood cells and immediately stored at-80C° until further processing.

- PE was defined as hypertension (blood pressure ≥140/90 mmHg) in two determinations, 4 hours apart
 associated with proteinuria (> 300 mg/24 h or ≥1+, according to a routine urinalysis) after 20 weeks of
 gestation in previously normotensive women [15]. In the absence of proteinuria, PE was characterized based
- on the new modified guidelines recommended by the task force on Hypertension in pregnancy [16]. In 2 out
 of 16 PE cases included in this study, the diagnosis was established on the presence of new onset
- 94 hypertension and impaired liver enzymes concentration.
- FGR was diagnosed when the estimated fetal weight, calculated using the Hadlock formula (Astraia
 Software GmbH), was below the 10th percentile for the evaluated gestational age [17].
- 97 Pregnant women with multiple pregnancies, congenital or chromosomal abnormalities of the fetus, renal
 98 disease, chronic hypertension, cardiovascular disease or other pregnancy complications were excluded from
 99 the study.
- The study was approved by the hospital's ethics committee according to the Helsinki Declaration on ethical
 principles for medical research involving human subjects. A written consent form was obtained from all
 participants prior to surgery.
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104 MicroRNA extraction and expression profiling

For total RNA extraction, 100mg of placental tissue were mechanically homogenized in TRIzol (Life
Technologies, Foster City, CA), treated with DNase I (Ambion, Austin, USA) and enriched for small RNAs
(siRNAs, miRNAs) using the mirVana microRNA Isolation kit (Ambion, Austin, USA). RNA quantity and
purity were measured using a Nanodrop spectrophotometer (ND-1000, NanoDrop Technologies, Houston,
TX, USA).

- 110 Labeling and hybridization were performed using the LabelIT miRNA labeling kit (Mirus Bio LLC, USA)
- according to the manufacturer's instructions. Samples were hybridized to an Applied MicroArrays (miRlink

112 Bioarray 300054-3PK) platform containing 1211 human miRNAs (Applied Biosystems, Foster City, CA).

113 This platform consists of 8 arrays per slide and on each array miRNAs are represented by multiple replicates.

- 114 Images were scanned using an Agilent Microarray Scanner (G2565CA).
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116 Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

117 The differential expression of miR-518a-5p in PE complicated placentas, as compared to the control group, was verified using Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR). Complementary DNA 118 (cDNA) synthesis and subsequent RT-PCR were performed, using the TaqMan miRNA Reverse 119 Transcription kit and TaqMan universal PCR master mix (Applied Biosystems, Inc., USA) respectively. 120 121 Specific primers and a TaqMan probe for miR-518a-5p (PN4427975, Applied Biosystems, Inc., USA) were 122 used according to the manufacturer's instructions. All samples were analyzed in triplicate in an LC480 LightCycler system (Roche GmbH, Switzerland). Positive amplification of each sample was considered 123 when a signal occurred before 40th threshold cycle. The miRNA levels were normalized to RNU44 (Applied 124 125 Biosystems, Inc., USA) as an internal control and the relative abundance of each miRNA was calculated 126 using the $\Delta \Delta Ct$ equation.

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128 Data analyses and statistics

129 Microarray Data Extraction and Analysis

130 Microarray raw data were extracted, as previously described, using the Imagene 6.0 software (Biodiscovery 131 Inc., USA) [18]. The MATLAB® simulation environment (The Mathworks, Inc., Natick, MA) was applied 132 for multi-parameter analyses. Background correction was performed by subtracting the median local background from the signal intensity. Normalization was performed using the quantile normalization 133 algorithm. The two-tailed student t-test was used to assess the mean differences between the two groups. 134 Relative expression was estimated as the *log*₂-transformed ratio of the individual miRNA expression levels 135 over the mean expression level of all control samples for the respective miRNA under investigation. Final 136 miRNA values have been calculated as the mean of each specific miRNA replicates on each array. 137 Continuous variables were expressed as median ± standard deviation, unless differently indicated. MiRNAs 138 139 were considered significant if they had a *p*-value<0.05 and False Discovery Rate (FDR)≤0.05. Microarray 140 platform as well as raw data have been uploaded on the Gene Expression Omnibus (GEO) database (https://www.ncbi.nlm.nih.gov/geo/) with reference numbers GPL23980 and GSE103542 respectively. 141

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143 Group-wise Comparison of miRNA expression and clinical data

MiRNA expression values were compared with clinical parameters, including PE (EOPE and LOPE), as well as the presence of proteinuria and FGR with the *Kruskal-Wallis* test. *Bonferroni* correction was used in order to adjust *p*-values for the statistical test performed and reduce the chances of obtaining false-positive results (type I errors) with the multiple pair-wise tests performed on the present dataset. Differences were considered significant at p < 0.02.

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151 *ROC Analysis*

152 Receiver Operating Characteristic (ROC) curves were performed with the MATLAB® simulation 153 environment (The Mathworks, Inc, Natick, MA) to evaluate the diagnostic potential of differentially 154 expressed miRNAs with 95% Standard Error (SE) and 95% Confidence Intervals (CI). ROC curves were 155 considered significant when an Area Under the Curve (AUC) value > 0.8 at a p < 0.05 was obtained.

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157 MiRNA Enrichment, Gene Ontology and Pathway Analysis

Differentially expressed miRNAs were further enriched and analyzed for known functions and pathway participation. Functional annotation was performed using Webgestalt web-tool [19]. RNA target prediction for selected miRNAs was performed with TargetScan v.7.0 [20], DIANA tools [21-23] and the miROB annotation web tool (<u>http://mirob.interactome.ru/</u>). Further on, we have used the GOmiR [24] multi-compare tool for a more in-depth functional annotation of investigated microRNAs.

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165 **RESULTS**

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167 MicroRNA expression profiles between PE and the control group

- 168 Overall, 44 miRNAs were dysregulated in PE-complicated placentas, as compared to the control cohort,
- 169 with a two-fold change and statistical significance (p < 0.05) (Figure 1). The vast majority was up-regulated
- 170 with a total of 27 miRNAs (61.4%) showing over-expression and 17 (38.6%) decreased expression.
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172 MicroRNA expression in early-onset and late-onset PE

- When comparing miRNA expression in samples obtained from women with EOPE (n=11) and LOPE (n=5)173 with controls (n=8), 9 miRNAs were found associated with the early or the late type of the disease (Figure 174 2). Specifically, miR-431 (p=0.003), miR-518a-5p (p=0.002) and miR-124* (p=0.001) were significantly 175 over-expressed in EOPE complicated placentas, as compared to controls, whereas miR-544b (p=0.001) was 176 significantly down-regulated and miR-3942 was marginally down-regulated (p=0.02) in EOPE when 177 compared to controls. Further on, miR-130b (p=0.004) and miR-423-3p (p=0.001) were significantly up-178 regulated in EOPE placentas as compared to controls, while miR-383 was marginally up-regulated in EOPE 179 (p=0.02) and significantly up-regulated (p=0.01) in LOPE samples. MiR-1183 was marginally up-regulated 180 (p=0.02) in LOPE samples, as compared to controls. In the EOPE placentas, following Bonferroni 181 correction, the over-expression of miR-1183 was not considered significant (p=0.03). 182
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184 MicroRNA expression in pregnancies complicated by FGR

- The miRNA expression profile was also compared in cases of PE and PE-FGR. Two miRNAs were found
 overexpressed in PE-FGR complicated placentas, including miR-431 and miR-518a-5p (Figure 3).
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188 **ROC analysis**

- ROC analysis revealed that 8 miRNAs up-regulated in PE complicated placentas (miR-500a, miR-383, miR-518a-5p, miR-431, miR-423-3p, miR-124*, miR-1183 and miR-130b) and 2 down-regulated (miR-3942, miR-544b) were associated with PE and miRNAs with p<0.05 and AUC>0.8 were considered as potential
- biomarkers. The ROC curves yielded the following AUCs: miR-500a (AUC=0.8), miR-3942 (AUC=0.8),
- 193 miR-544b (AUC=0.84), miR-383 (AUC≈0.82), miR-518a-5p (AUC=0.83), miR-431 (AUC=0.82), miR-423-
- 194 3p (AUC≈0.90), miR-124* (AUC=0.88), miR-1183 (AUC=0.80) and miR-130b (AUC=0.82) (Figure 4).
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196 **qRT PCR verification**

The over-expression of miR-518a-5p in PE complicated placentas as compared to controls was verified using
 qRT-PCR in all samples. Consistent with microarray results the change of dysregulated miRNA was
 approximately two fold above (Figure 1 suppl.).

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203 MiRNA Enrichment, Gene Ontology and Pathway Analysis

204 Several genes have been reported as predicted or verified targets of the aforementioned miRNAs. 205 Specifically, miR-126*, miR-885, miR-130B, miR-1915, miR-1204, miR-3928, miR-518a-5p, miR-631, 206 miR-383, miR-19a, miR-302c, miR-155, miR-30a, miR-542 and miR-1248 manifested significant associations with genes including SMAD1/2/4/5, ADAM9, VCAM1, TGFBP2, PTEN, TP53 and RUNX2 207 208 (Figure 2 suppl.). Pathway analysis of significantly annotated miRNAs and their respective targets revealed that they participate in biological pathways associated with PE including MAPK, TNF signaling, T-cell 209 receptor, B-cell receptor and TGF- β pathways (Figure 5). Further on, the GOmiR tool searches in the 210 MiRanda, TargetScan, PicTar, Sanger, RNAhybrid databases unraveled common gene and functional targets 211 for all the aforementioned miRNAs. In particular, GOmiR confirmed the results obtained by the other tools, 212 revealing that miR-126* and miR-130B target genes participate in TNF-signaling. It also revealed PPARG, 213 another target gene, which participates in obstetric diseases such as Polycystic Ovary Syndrome, Ovarian 214 Cysts, Ovarian Diseases and Metabolic Syndrome. In addition, GOmiR confirmed that miR-518a-5p 215 participates in MAPK signaling. Further on, miR-631 was found to target TBX2, HDAC1 and AXIN1 which 216 217 participate in ectodermal placode formation. Target genes of the miR-631 were shown to participate in metabolic pathways and the insulin signaling pathway, whereas miR-30a target genes are involved in 218 219 MAPK, JAK-STAT signaling pathways as well as in metabolic pathways. 220

222 DISCUSSION

Several studies have demonstrated dysregulation of placenta miRNAs in PE, implying their involvement in the pathogenesis of the disease by targeting a variety of genes and regulating crucial biological pathways [5-7, 10, 11, 14, 25, 26]. Despite extensive efforts, however, conflicting results exist which may be attributed to the small number of samples included in each study, the increased number of molecules analyzed, the position of the placenta from which samples are obtained, the gestational age at sampling and the differences in the platforms used for analysis [7].

verified the differential expression of several miRNAs previously reported, including the over-expression of
 miR-155 and miR-130b, as well as the down-regulation of miR-126* [11,25, 27-35].

MiR-155 and miR-130b have been implicated in the pathogenesis of PE through their involvement in pathways regulating trophoblast cells such as TGF- β signaling [11, 27-30]. Recently, miR130b was reported as differentially expressed in relation to pre-pregnancy BMI and adipogenesis, both of which have been associated with increased risk for the development of pregnancy complications including PE [31-33].

236 Although conflicting results exist regarding the role of mir-126 in the pathogenesis of PE, miR-126 and its isoform 126* regulate many target genes mainly related to angiogenesis [25, 34, 35]. Notably, in vitro 237 experiments, using endothelial cells, revealed that miR-126 positively regulates their migration and 238 239 proliferation, by enhancing fibroblast growth during arterial development [35]. Lack of miR126 has a 240 negative angiogenic response, through VEGFA, bFGF and MAP kinase [34, 36]. In the current study, in 241 silico analysis also revealed that miR126* can potentially modulate ADAM9, MMP7, MMP13, VEGFA and 242 VCAM1. Proteins encoded by these genes are currently under investigation as potential biomarkers for the identification of pregnant women destined to develop PE. 243

Over-expression of miR-423-3p, miR-124*, miR-500a, miR-431 and miR-1305 and down-regulation of 244 miR-544 and miR-3942 were observed in PE complicated placentas, as compared to controls. These 245 246 miRNAs have not been previously associated with the pathogenesis of PE. MiR-423-3p, has been linked to renal function, acute kidney injury and cell proliferation [37, 38]. MiR-124 has been implicated in 247 angiogenesis through the renin-angiotensin system and is hence linked with the regulation of blood pressure 248 249 [39]. MiR-1305 has been recently implicated in cell cycle regulation and apoptosis in pluripotent stem cells by targeting CDK6, CYCLIND2 and RUNX2 [40]. Mir-431, a member of C14MC placenta cluster, directly 250 251 regulates the expression of SMAD4 [41]. Over-expression may negatively regulate IGF1R and consequently inhibit cell proliferation through MAPK pathway, which plays an important role in the development of 252 253 vascular lesions that are observed in diabetes and hypertension [42].

Results were also analyzed with respect to the gestational age of disease onset. In the literature, a limited number of studies have focused on comparisons between miRNA expression pattern in EOPE and LOPE. In the current study, when correlations between EOPE, LOPE and the control cohort were performed, 9 miRNAs were found aberrantly expressed. Specifically, miR-383 and miR-1183 were over-expressed in PE, higher in LOPE cases as compared to controls, possibly reflecting an association with a milder form of the disease. Additionally, higher levels of expression for miR-518a-5p, miR-423, miR-124* and miR-431 were

- detected in EOPE as compared to controls. It seems likely that all four miRNAs are associated with the severity of the disease and the severity of the clinical symptoms. On the contrary, miR-544 and miR-3942 were down-regulated in the EOPE cases, indicating a putative PE-specific diagnostic marker. Mir-130b was found over-expressed in the EOPE cases when compared to the LOPE, signifying its potential use as a biomarker for severe PE.
- 265 When examining miRNA expression profiles in PE complicated by FGR, miR-518-5p and miR-431 were up-
- regulated in all PE-FGR cases, indicating their potential as unfavorable prognostic markers. It is noticeable
- that overexpression of both miRNAs was also observed in previous correlations, reinforcing the prediction
- about their possible linkage to poor PE outcomes when up-regulated.
- Although the properties of miR-518a-5p remain to be elucidated, statistically significant over-expression was found in all associations performed in the present study, including PE diagnosis, EOPE, LOPE and PE-FGR following ROC analysis. Vashukova *et al.*, reported up-regulation of miR-518a-5p in superimposed PE on chronic hypertension [43]. The aforementioned observation, along with the data obtained in the present study, possibly indicate a common pathway in the pathogenesis of EOPE, superimposed PE and PE-FGR. Since miR-518a-5p is also expressed in maternal peripheral blood during pregnancy, it represents a potential noninvasive diagnostic biomarker for all types of PE [10]. These findings however, should be validated in
- 276 larger PE cohorts.
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279 **REFERENCES**

- [1] U.D. Anderson, M.G. Olsson, K.H. Kristensen, B. Akerstrom, S.R. Hansson, Review: Biochemical
 markers to predict preeclampsia, Placenta 33 Suppl (2012) S42-7.
- [2] L. Myatt, J.M. Roberts, Preeclampsia: Syndrome or Disease?, Current hypertension reports 17(11) (2015)
 83.
- [3] M. Ha, V.N. Kim, Regulation of microRNA biogenesis, Nat Rev Mol Cell Biol 15(8) (2014) 509-24.
- [4] A. Lycoudi, D. Mavreli, A. Mavrou, N. Papantoniou, A. Kolialexi, miRNAs in pregnancy-related
 complications, Expert review of molecular diagnostics 15(8) (2015) 999-1010.
- [5] B.L. Pineles, R. Romero, D. Montenegro, A.L. Tarca, Y.M. Han, Y.M. Kim, S. Draghici, J. Espinoza,
 J.P. Kusanovic, P. Mittal, S.S. Hassan, C.J. Kim, Distinct subsets of microRNAs are expressed differentially
 in the human placentas of patients with preeclampsia, American journal of obstetrics and gynecology 196(3)
 (2007) 261 e1-6.
- [6] Y. Hu, P. Li, S. Hao, L. Liu, J. Zhao, Y. Hou, Differential expression of microRNAs in the placentae of
 Chinese patients with severe pre-eclampsia, Clinical chemistry and laboratory medicine 47(8) (2009) 923-9.
- [7] P. Xu, Y. Zhao, M. Liu, Y. Wang, H. Wang, Y.X. Li, X. Zhu, Y. Yao, H. Wang, J. Qiao, L. Ji, Y.L.
 Wang, Variations of microRNAs in human placentas and plasma from preeclamptic pregnancy,
 Hypertension 63(6) (2014) 1276-84.
- [8] A.M. Sheikh, H.Y. Small, G. Currie, C. Delles, Systematic Review of Micro-RNA Expression in PreEclampsia Identifies a Number of Common Pathways Associated with the Disease, PloS one 11(8) (2016)
 e0160808.
- [9] D.A. Enquobahrie, D.F. Abetew, T.K. Sorensen, D. Willoughby, K. Chidambaram, M.A. Williams,
 Placental microRNA expression in pregnancies complicated by preeclampsia, American journal of obstetrics
 and gynecology 204(2) (2011) 178 e12-21.
- I. Hromadnikova, L. Dvorakova, K. Kotlabova, A. Kestlerova, L. Hympanova, V. Novotna, J. Doucha,
 L. Krofta, Circulating heat shock protein mRNA profile in gestational hypertension, pre-eclampsia & foetal
 growth restriction, The Indian journal of medical research 144(2) (2016) 229-237.
- [11] X. Li, C. Li, X. Dong, W. Gou, MicroRNA-155 inhibits migration of trophoblast cells and contributes to
 the pathogenesis of severe preeclampsia by regulating endothelial nitric oxide synthase, Mol Med Rep 10(1)
 (2014) 550-4.
- [12] M. Zhang, S. Muralimanoharan, A.C. Wortman, C.R. Mendelson, Primate-specific miR-515 family
 members inhibit key genes in human trophoblast differentiation and are upregulated in preeclampsia,
 Proceedings of the National Academy of Sciences of the United States of America (2016).
- [13] L. Xie, Y. Sadovsky, The function of miR-519d in cell migration, invasion, and proliferation suggests a
 role in early placentation, Placenta 48 (2016) 34-37.
- [14] T. Gunel, M.K. Hosseini, E. Gumusoglu, H.I. Kisakesen, A. Benian, K. Aydinli, Expression profiling of
 maternal plasma and placenta microRNAs in preeclamptic pregnancies by microarray technology, Placenta
 52 (2017) 77-85.
- [15] C.M. Brown, V.D. Garovic, Mechanisms and management of hypertension in pregnant women, Current
 hypertension reports 13(5) (2011) 338-46.
- [16] O. American College of, Gynecologists, P. Task Force on Hypertension in, Hypertension in pregnancy.
 Report of the American College of Obstetricians and Gynecologists' Task Force on Hypertension in
- 320 Pregnancy, Obstetrics and gynecology 122(5) (2013) 1122-31.

- [17] O. American College of, Gynecologists, ACOG Practice bulletin no. 134: fetal growth restriction,
 Obstetrics and gynecology 121(5) (2013) 1122-33.
- [18] M. Braoudaki, G.I. Lambrou, K. Giannikou, V. Milionis, K. Stefanaki, D.K. Birks, N. Prodromou, A.
 Kolialexi, A. Kattamis, C.A. Spiliopoulou, F. Tzortzatou-Stathopoulou, E. Kanavakis, Microrna expression
 signatures predict patient progression and disease outcome in pediatric embryonal central nervous system
 neoplasms, J Hematol Oncol 7 (2014) 96.
- [19] B. Zhang, S. Kirov, J. Snoddy, WebGestalt: an integrated system for exploring gene sets in various
 biological contexts, Nucleic Acids Res 33(Web Server issue) (2005) W741-8.
- [20] V. Agarwal, G.W. Bell, J.W. Nam, D.P. Bartel, Predicting effective microRNA target sites in
 mammalian mRNAs, Elife 4 (2015).
- [21] M.D. Paraskevopoulou, G. Georgakilas, N. Kostoulas, I.S. Vlachos, T. Vergoulis, M. Reczko, C.
 Filippidis, T. Dalamagas, A.G. Hatzigeorgiou, DIANA-microT web server v5.0: service integration into
 miRNA functional analysis workflows, Nucleic acids research 41(Web Server issue) (2013) W169-73.
- [22] M.D. Paraskevopoulou, G. Georgakilas, N. Kostoulas, M. Reczko, M. Maragkakis, T.M. Dalamagas,
 A.G. Hatzigeorgiou, DIANA-LncBase: experimentally verified and computationally predicted microRNA
 targets on long non-coding RNAs, Nucleic acids research 41(Database issue) (2013) D239-45.
- [23] I.S. Vlachos, N. Kostoulas, T. Vergoulis, G. Georgakilas, M. Reczko, M. Maragkakis, M.D.
 Paraskevopoulou, K. Prionidis, T. Dalamagas, A.G. Hatzigeorgiou, DIANA miRPath v.2.0: investigating the
 combinatorial effect of microRNAs in pathways, Nucleic acids research 40(Web Server issue) (2012) W498 504.
- [24] M.G. Roubelakis, P. Zotos, G. Papachristoudis, I. Michalopoulos, K.I. Pappa, N.P. Anagnou, S.
 Kossida, Human microRNA target analysis and gene ontology clustering by GOmir, a novel stand-alone
 application, BMC bioinformatics 10 Suppl 6 (2009) S20.
- [25] O. Ishibashi, A. Ohkuchi, M.M. Ali, R. Kurashina, S.S. Luo, T. Ishikawa, T. Takizawa, C. Hirashima,
 K. Takahashi, M. Migita, G. Ishikawa, K. Yoneyama, H. Asakura, A. Izumi, S. Matsubara, T. Takeshita, T.
 Takizawa, Hydroxysteroid (17-beta) dehydrogenase 1 is dysregulated by miR-210 and miR-518c that are
 aberrantly expressed in preeclamptic placentas: a novel marker for predicting preeclampsia, Hypertension
 59(2) (2012) 265-73.
- [26] L. Jiang, A. Long, L. Tan, M. Hong, J. Wu, L. Cai, Q. Li, Elevated microRNA-520g in pre-eclampsia
 inhibits migration and invasion of trophoblasts, Placenta 51 (2017) 70-75.
- [27] Y. Dai, Z. Qiu, Z. Diao, L. Shen, P. Xue, H. Sun, Y. Hu, MicroRNA-155 inhibits proliferation and
 migration of human extravillous trophoblast derived HTR-8/SVneo cells via down-regulating cyclin D1,
 Placenta 33(10) (2012) 824-9.
- [28] Y. Zhang, Z. Diao, L. Su, H. Sun, R. Li, H. Cui, Y. Hu, MicroRNA-155 contributes to preeclampsia by
 down-regulating CYR61, Am J Obstet Gynecol 202(5) (2010) 466 e1-7.
- [29] W. Cheng, T. Liu, F. Jiang, C. Liu, X. Zhao, Y. Gao, H. Wang, Z. Liu, microRNA-155 regulates
 angiotensin II type 1 receptor expression in umbilical vein endothelial cells from severely pre-eclamptic
 pregnant women, Int J Mol Med 27(3) (2011) 393-9.
- [30] L. Wu, H. Zhou, H. Lin, J. Qi, C. Zhu, Z. Gao, H. Wang, Circulating microRNAs are elevated in plasma
 from severe preeclamptic pregnancies, Reproduction 143(3) (2012) 389-97.
- [31] D.A. Enquobahrie, M. Hensley, C. Qiu, D.F. Abetew, K. Hevner, M.G. Tadesse, M.A. Williams,
 Candidate Gene and MicroRNA Expression in Fetal Membranes and Preterm Delivery Risk, Reproductive
 sciences 23(6) (2016) 731-7.

- 364 [32] Z. Chen, J. Luo, L. Ma, H. Wang, W. Cao, H. Xu, J. Zhu, Y. Sun, J. Li, D. Yao, K. Kang, D. Gou,
- 365 MiR130b-Regulation of PPARgamma Coactivator- 1alpha Suppresses Fat Metabolism in Goat Mammary
- 366 Epithelial Cells, PloS one 10(11) (2015) e0142809.
- [33] D.A. Enquobahrie, P.L. Wander, M.G. Tadesse, C. Qiu, C. Holzman, M.A. Williams, Maternal pre pregnancy body mass index and circulating microRNAs in pregnancy, Obesity research & clinical practice
 (2016).
- [34] S. Wang, A.B. Aurora, B.A. Johnson, X. Qi, J. McAnally, J.A. Hill, J.A. Richardson, R. Bassel-Duby,
 E.N. Olson, The endothelial-specific microRNA miR-126 governs vascular integrity and angiogenesis,
 Developmental cell 15(2) (2008) 261-71.
- [35] J.E. Fish, M.M. Santoro, S.U. Morton, S. Yu, R.F. Yeh, J.D. Wythe, K.N. Ivey, B.G. Bruneau, D.Y.
 Stainier, D. Srivastava, miR-126 regulates angiogenic signaling and vascular integrity, Developmental cell
 15(2) (2008) 272-84.
- [36] F. Kuhnert, M.R. Mancuso, J. Hampton, K. Stankunas, T. Asano, C.Z. Chen, C.J. Kuo, Attribution of
 vascular phenotypes of the murine Egfl7 locus to the microRNA miR-126, Development 135(24) (2008)
 3989-93.
- [37] H.T. Li, H. Zhang, Y. Chen, X.F. Liu, J. Qian, MiR-423-3p enhances cell growth through inhibition of
 p21Cip1/Waf1 in colorectal cancer, Cell Physiol Biochem 37(3) (2015) 1044-54.
- [38] N. Bruno, J.M. ter Maaten, E.S. Ovchinnikova, E.L. Vegter, M.A. Valente, P. van der Meer, R.A. de
 Boer, P. van der Harst, D. Schmitter, M. Metra, C.M. O'Connor, P. Ponikowski, J.R. Teerlink, G. Cotter, B.
 Davison, J.G. Cleland, M.M. Givertz, D.M. Bloomfield, H.C. Dittrich, Y.M. Pinto, D.J. van Veldhuisen,
 H.L. Hillege, E. Berezikov, A.A. Voors, MicroRNAs relate to early worsening of renal function in patients
 with acute heart failure, Int J Cardiol 203 (2016) 564-9.
- [39] S. Sober, M. Laan, T. Annilo, MicroRNAs miR-124 and miR-135a are potential regulators of the
 mineralocorticoid receptor gene (NR3C2) expression, Biochemical and biophysical research
 communications 391(1) (2010) 727-32.
- [40] S. Jin, J. Collin, L. Zhu, D. Montaner, L. Armstrong, I. Neganova, M. Lako, A Novel Role for miR1305 in Regulation of Pluripotency-Differentiation Balance, Cell Cycle, and Apoptosis in Human Pluripotent
 Stem Cells, Stem Cells 34(9) (2016) 2306-17.
- [41] K.P. Lee, Y.J. Shin, A.C. Panda, K. Abdelmohsen, J.Y. Kim, S.M. Lee, Y.J. Bahn, J.Y. Choi, E.S.
 Kwon, S.J. Baek, S.Y. Kim, M. Gorospe, K.S. Kwon, miR-431 promotes differentiation and regeneration of
 old skeletal muscle by targeting Smad4, Genes Dev 29(15) (2015) 1605-17.
- [42] T. Tanaka, S. Sugaya, K. Kita, M. Arai, T. Kanda, K. Fujii, F. Imazeki, K. Sugita, O. Yokosuka, N.
 Suzuki, Inhibition of cell viability by human IFN-beta is mediated by microRNA-431, Int J Oncol 40(5)
 (2012) 1470-6.
- [43] E.S. Vashukova, A.S. Glotov, P.V. Fedotov, O.A. Efimova, V.S. Pakin, E.V. Mozgovaya, A.A.
 Pendina, A.V. Tikhonov, A.S. Koltsova, V.S. Baranov, Placental microRNA expression in pregnancies
 complicated by superimposed preeclampsia on chronic hypertension, Mol Med Rep 14(1) (2016) 22-32.
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403 FIGURE LEGENDS

Figure 1. Histogram of the 44 dysregulated miRNAs in PE complicated placentas as compared to controls (p < 0.05 and FDR ≤ 0.05). Over-expressed miRNAs are indicated with upward bars and down-regulated with downward bars. Y axis values represent miRNA fold change (ratios presented include Early Onset Preeclampsia (EOPE) (n=11) and Late Onset Preeclampsia (LOPE) (n=5).

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Figure 2. Differences in miRNA expression in EOPE, LOPE and control placentas. Significant differential expression was identified for (A) miR-3942 between EOPE and controls (p=0.02), (B) miR-544b between EOPE and controls (p=0.001), (C) miR-383 between EOPE and controls as well as LOPE and controls (p=0.02and p=0.01 respectively), (D) miR-518a-5p between EOPE and controls (p=0.002), (E) miR-431 between EOPE and controls (p=0.003), (F) miR-423-3p between EOPE and controls as well as LOPE and controls (p=0.001 and p=0.007 respectively), (G) miR-124* between EOPE and controls (p=0.001), (H) miR-1183 between EOPE and controls as well as LOPE and controls (p=0.03 and p=0.02 respectively) and (I) miR-130b between EOPE and controls as well as LOPE and controls (p=0.004 and 0.04 respectively) (EOPE: Early Onset Preeclampsia, LOPE: Late Onset Preeclampsia) (Expression levels concern the natural, log_2 untransformed miRNA expression values. * depicts a significance at the p<0.05 level and ** depicts a significance at the p<0.01 level).

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Figure 3. Over-expression of (A) miR-518a-5p and (B) miR-431 in PE complicated placentas with FGR fetuses as compared to those with unaffected fetal growth (PE: Preeclampsia, FGR: Fetal Growth Restriction) (Expression levels correspond to the natural log_2 untransformed expression values. Differences were significant at p<0.05).

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Figure 4. Receiver Operating Characteristic (ROC) curves of miRNAs differentiating controls from Preeclampsia complicated placentas. Ten miRNAs can differentiate controls from Preeclampsia complicated placentas: (A) miR-500a, (B) miR-03942, (C) miR-544b, (D) miR-383, (E) miR-518a-5p, (F) miR-431, (G) miR-423-3p, (H) miR-124*, (I) miR-1183 and (J) miR-130b manifested an Area Under the Curve (AUC) value greater than 0.8 (AUC>0.8) and p<0.05, suggesting that they are possible candidates for Preeclampsia.

424 Figure 5. Selected significant signaling pathways associated with annotated differentially expressed miRNAs (miR-125b, miR-155-5p, miR-21-5p and miR-145).

425 MiRNA annotation was performed with the DIANA mIRNA web annotation tool <u>http://diana.imis.athena-innovation.gr/DianaTools/index.php?r=mirpath/index</u>).

Table 1. Maternal and neonatal characteristics of Preeclampsia cases and controls included in the study.										
	Controls (n=8)	EOPE* (n=11)	LOPE** (n=5)	p-value ¹	p-value ²	p-value ³	p-value ⁴			
Maternal Age (y)	35.7 [35-39]	35.1 [28-45]	28.4 [20-35]	0.051	0.820	0.036	0.055			
Pre-pregnancy BMI (kg/m2)	29.2 [22.96-31.99]	29.9 [19.83-41.77]	28.6[24.02 - 36.57]	0.942	0.835	0.867	0.778			
Blood Pressure (mmHg)										
Systolic	121 [122-118]	197.75 [160-250]	172.25 [152-197]	0.702	0.889	0.612	0.412			
Diastolic	75 [72-85]	113.5 [90-150]	107.75 [94-135]	0.966	0.962	0.923	0.798			
Proteinuria (n)										
Yes	0	10 (91%)	4 (80%)	$0.0002^{\text{¥}}$						
No	8 (100%)	1 (9%)	1 (20%)	0.0002						
Fetal Growth Restriction (FGR)	None	6 (54.5%)	2 (40%)							
Gestational Age at delivery (wk)	36.3 [34.2 - 38.5]	31.2 [28 - 33.5]	35.6 [34.4 - 37.3]	0.051	0.0003	0.29	0.78			
Fetal Birth Weight (grams)	2792.6 [2215 - 3560]	1335 [1025 - 1540]	2454 [1900 - 3020]	0.0003	0.0002	0.336	0.002			
>10th percentile (n)	8 (100%)	5 (45.4%)	3 (60%)							
<10th percentile (n)	0	4 (36.4%)	1 (20%)							
<5th percentile (n)	0	2 (18.2%)	1 (20%)							

*EOPE: Early onset Preeclampsia ** LOPE: Late onset Preeclampsia

¹Significance level between all groups

²Significance level between Control and EOPE placentas

³Significance level between Control and LOPE placentas

⁴Significance level between EOPE and LOPE placentas

[¥]p-value estimated with *chi-square* test for independence between proteinuria and early and late onset Preeclampsia.

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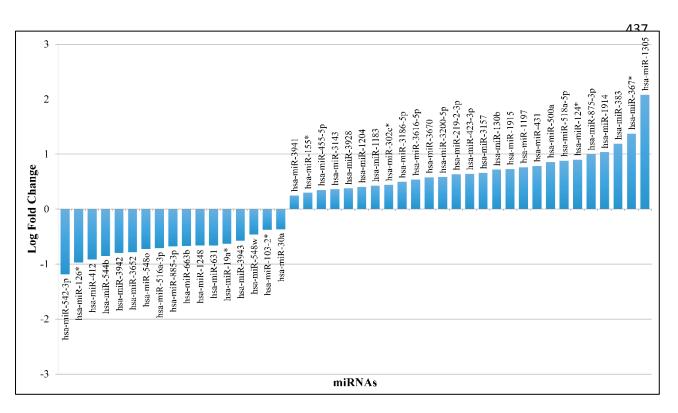
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435 Figure 1.

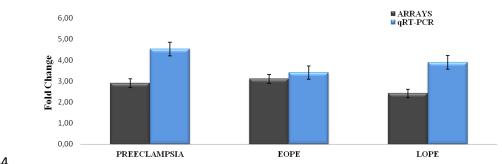




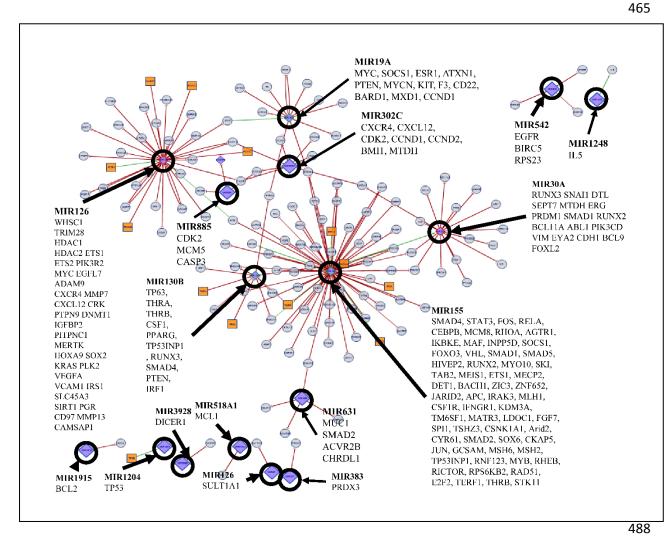
461 SUPPLEMENTARY FIGURES

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463 Supplementary Figure 1.



Comparative diagram of miRNA expression between qRT-PCR and microarrays



Supplementary Figure 2. Networks of miRNAs and their association with target genes. In particular, we have found that miR-126, miR-885, miR-130b, miR-1915, miR-1204, miR-3928, miR-518a1, miR-126, miR-631, miR-383, miR-19a, miR-302c, miR-155, miR-30a, miR-542, miR-1248, manifested significant associations with gene groups. miRNA annotation was performed with the miROB web tool (http://mirob.interactome.ru/).